

Elite™ gp120 (HIV-1/Clade B) ELISA Assay Kit

CATALOG NUMBER: CA-gp120B, 200 assays

Description

gp120 (HIV-1/Clade B) ELISA Assay Kit contains the key components required for the quantitative analysis of gp120 (HIV-1/Clade B) concentrations in cell culture supernatants and serum within the range of 0.316-316 ng/ml in a sandwich ELISA format. A pair of monoclonal antibody and polyclonal antibody has been selected as capture antibody and detection antibody. The components supplied in this kit are sufficient to assay gp120 (HIV-1/Clade B) in two 96-well ELISA plates.

Kit Components

- Component A: Capture Antibody (anti-gp120 (HIV-1/Clade B) monoclonal antibody)
 50 μl (1 mg/ml)
- Component B: gp120 Standard (recombinant gp120 (HIV-1/Clade B) protein)
 25 μI (50 μg/ml)
- Component C: Detection Antibody (anti-gp120 (HIV-1/Clade B) rabbit polyclonal antibody)
- Component D: HRP-Conjugated Anti-Rabbit IgG 25 μI
- Component E: TMB Solution A (3,3',5,5'- tetramethylbenzidine) (light sensitive!)
- Component F: TMB Solution B (H₂O₂) (light sensitive!)
 12 ml
- Component G: TMB Stop Solution 25 ml

Materials Required but not Provided

- 96-well microtiter plates for ELISA assay
- PBS
- Coating Buffer: 0.05 M Carbonate-Biocarbonate, pH 9.6
- Blocking Buffer: 5% milk in PBS
- Washing Buffer: 0.05% Tween-20 in PBS
- Dilution Solution: 0.05% Tween-20 and 0.5% milk in PBS

Storage

Keep in 4 °C and avoid exposure to light; do not freeze!

Assay Protocol

1. Plate Preparation:

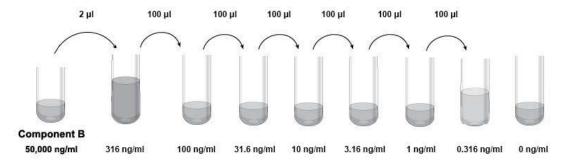
- 1.1 For each 96-well microtiter plate, dilute 20 μl of Capture Antibody (**Component A**) with 10.5 ml of Coating Buffer to prepare a coating solution. Immediately add 100 μl of the coating solution to each well. Seal the plate and incubate overnight at 4 °C.
- 1.2 Remove the coating solution by decanting. Invert the plate and blot it briefly with clean paper towel.
- 1.3 Add 300 µl of Blocking Buffer to each well. Incubate for at least 2 hours at 37 °C.
- 1.4 Aspirate to remove Blocking Buffer and wash the plate 4 times with 300 µl of Washing Buffer per well.

2. Run ELISA Assay:





2.1 Standard/Sample: Dilute the gp120 Standard (**Component B**) with PBS to eight concentrations (316 ng/ml , 100 ng/ml, 31.6 ng/ml, 10 ng/ml, 3.16 ng/ml, 1 ng/ml, 0.316 ng/ml, and 0 ng/ml). Immediately add 100µl of Standard and sample to each well in triplicate. Incubate at 37 °C for 1 hour.



- 2.2 Detection: Aspirate and wash plate 4 times. Dilute 10 μl of Detection Antibody (**Component C**) with 10.5 ml of Dilution Solution to prepare a detection solution. Add 100 μl of the detection solution into each well. Incubate at 37 °C for 1 hour.
- 2.3 Anti-Rabbit IgG Peroxidase: Aspirate and wash plate 4 times. Dilute 10 µl of HRP-Conjugate Anti-Rabbit IgG (**Component D**) with 10.5 ml of Dilution Solution. Add 100 µl into each well. Incubate at 37 °C for 1 hour.
- 2.4 TMB Peroxidase Substrate Solution Preparation: Mix equal volumes of TMB Solution A (**Component E**) and TMB Solution B (**Component F**) in a clean, preferably HDPE, polypropylene or glass container immediately prior to use at room temperature.
 - Note: For one 96-well plate, prepare 12 ml TMB Peroxidase Substrate Solution by mixing 6 ml **Component E** and **Component F**.
- 2.5 TMB Reaction: Aspirate and wash plate 4 times with Washing Buffer. Add 100 µl of TMB Peroxidase Substrate Solution (from Step 2.4) into each well. Incubate at 37 °C for 30 minutes.
- 2.6 TMB Reaction Termination: Add 100 µl of TMB Stop Solution (**Component G**) to each well. This stop solution will halt color development and will turn the TMB Substrate from blue into yellow.
- 2.7 Read: Determine the optical density of each well within 30 minutes using a microplate reader set to 450nm.
- 2.8 Analysis: Average the triplicate reading for each standard, control, and sample, then subtract the average zero standard optical density. Create a standard curve by reducing the data using computer software capable of generating a four parameter logistic (4-PL) or other curve-fit. The gp120 (HIV-1/Clade B) concentration in sample can be determined by regression analysis. If samples have been diluted, the concentration calculated from the standard curve must be multiplied by the dilution factor.

