Human Lysophosphatidic Acid Receptor 2 Stable Cell Line

Catalog #: CCL-LPAR2-4SH

Description: Tetracycline inducible recombinant human Lysophosphatidic acid receptor 2 stable CHO cell line

Provided Material

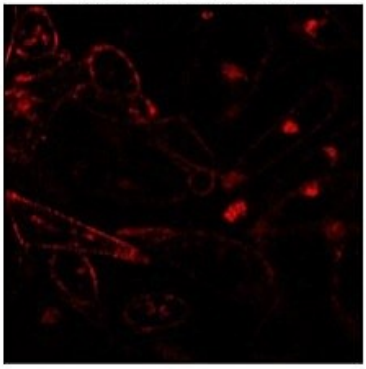
Cells	2 x 1 mL frozen aliquots
Format	3.10^6 cells/mL in freezing medium
Shipping conditions	Shipped on dry ice. Please ensure dry ice is still present in the package upon receipt
Storage condition	Store in liquid nitrogen (vapor phase) immediatly upon receipt

Product Informations

Uniprot accession	Q9HBW0
Cell line name	H-LPAR2
Receptor construct	**2*Twin-tag – TEV – LPAR2 – TEV – His**
Product description	Human lysophosphatidic acid receptor 2 stable CHO-TREx cell line was established using Hygromycin at 150µg/mL. This monoclonal cell line were obtained by limit dilution allowing a homogeneous expression, assesed by confocal immuno-imaging
Gene synonyms	EDG4, LPA2
Receptor synonyms	LPA receptor 2, LPA-2, Lysophosphatidic acid receptor Edg-4

Quality control

Immuno staining : *Strep-Tactin Chromeo 546*



Stability	The cells were kept in continuous culture for at least 60 days and showed no decrease of receptor expression level
Mycoplasma	This cell line was tested negative for mycoplasma *(MycoAlert™ mycoplasma detection kit – Lonza)*

Recommended culture conditions

Thawing protocol

Remove the vial from liquid nitrogen tank and place it quickly in a 37 °C water-bath.
Just before the cells are completely thawed, decontaminate the outside of the vial with 70% ethanol and transfer the cells to a 15mL centrifuge tube containing 9mL of complete growth medium.
Pellet cells by centrifugation at 200 x g for 5 min, and discard the supernatant.
Resuspend the cells pellet with 5mL of the recommended complete growth medium and dispense into a 75cm² culture flask (T-75) containing 10mL of the same medium.
Incubate the culture at 37°C in a humidified atmosphere with 5% CO_2.

Grow medium	*DMEM F-12, 10% FBS, 7µg/mL Blasticidin, 150µg/mL Hygromycin*
Freeze medium	*50% FBS, 40% DMEM F-12 and 10% DMSO*

Subculturing protocol

Remove and discard culture medium.
Wash cells with PBS (pH=7.4) to remove all traces of serum that contains trypsin inhibitor.
Add 3mL of 0.05% (w/v) Trypsin-EDTA solution to T-75 and observe the cells under an inverted microscope until cell layer is dispersed (usually 1 to 3 minutes).
Add 10 mL of complete growth medium and aspirate cells by gently pipetting, centrifuge the cells at 200 x g for 5min, and discard the supernatant.
Resuspend the cells in complete growth medium and add appropriate aliquots of the cell suspension to new culture vessels. A subcultivation ratio of 1:3 to 1:8 is recommended.
Incubate cultures at 37 °C in a humidified atmosphere with 5% CO_2.

Induction protocol

Notes: Please split the cells at least 2 times after thawing them before starting the induction protocol. For optimal induction, cells must reach 70-80% confluence.
Remove and discard culture medium.
Add an adequate volume of complete growth medium containing 2.5 µg/mL of Tetracycline.
Incubate the cells at 37°C in a humidified atmosphere with 5% CO_2 during 36h.
Before use check cells morphology.

Materials

Product Name	Provider	Cat.Ref
DMEM F-12	Gibco	31331
FBS	Gibco	10270
Trypsin – EDTA (0.05%)	Gibco	25300
DPBS (without Ca2+, Without Mg2+)	Gibco	14190
Blasticidin	InvivoGen	ant-bl-1
Hygromycin	InvivoGen	ant-hg-1
Tetracycline	Sigma	T7660
DMSO	Sigma	D4540
Strep-Tactin Chromeo 488	iba	2-1542-050