

Elite™ Caspase Activity Assay Kit (Green Fluorescence)

CATALOG NUMBER: CA-C061, 100 assays

Description

Caspases play important roles in apoptosis and cell signaling. The activation of caspase is widely accepted as a reliable indicator for cell apoptosis. Most caspases have substrate selectivity for the peptide sequence Val-Ala-Asp (VAD). This kit uses TF2-VAD-FMK as a fluorescent indicator for most caspase activities. The cell permeable and nontoxic TF2-VAD-FMK irreversibly binds to activated caspase-1, -3, -4, -5, -6, -7, -8 and -9 in apoptotic cells. Once bound to caspases, the fluorescent reagent is retained inside the cell. The binding event prevents the caspases from further catalysis but will not stop apoptosis from proceeding. Within 15-minute incubation, it starts to react with active caspase enzymes.

Elite™ Caspase Activity Assay Kit provides all the essential components with an optimized assay protocol. It is designed to detect cell apoptosis by measuring generic activation of caspases (caspase-1, -3, -4, -5, -6, -7, -8 and -9) in live cells. The kit is used for either the quantification of most activated caspase activities in apoptotic cells or screening of caspase inhibitors. TF2-VAD-FMK, the green label reagent, allows for direct detection of activated caspases in apoptotic cells by a flow cytometer at Ex/Em = 488/520 nm.

Features

- **Optimized Performance:** Optimal conditions for the detection of caspase activities.
- **Optimized Performance:** optimal for the detection of many caspase activities.
- **Convenient:** Formulated to have minimal hands-on time.
- **Non-radioactive:** No special requirement for waste disposal.

Kit Components

- | | |
|--------------------------------------|-------------------|
| • Component A: 500x TF2-VAD-FMK | 1 vial (100 µl) |
| • Component B: Assay Buffer | 50 ml in a bottle |
| • Component C: 500x Propidium Iodide | 1 vial (100 µl) |

Storage

Keep in freezer (-20 °C) and avoid exposure to light.

Materials Required (but not supplied)

- 96 or 384-well microplates: Tissue culture microplate with black wall and clear bottom is recommended.
- A fluorescence microplate reader: Capable of monitoring fluorescence intensity at Ex/Em = 490±10/520±10 nm.

Assay Protocol for 96-Well Plate

1. For each sample, prepare cells in 0.5 ml warm medium or buffer of your choice at a density of 5×10^5 to 1×10^6 cells/ml.
Note: Each cell line should be evaluated on an individual basis to determine the optimal cell density for apoptosis induction.
2. Treat cells with test compounds for a desired period to induce apoptosis, and create positive and negative controls
3. Add 1 µl of 500X TF2-VAD-FMK (Component A) into the treated cells (from Step 2), and incubate the cells in a 37 °C, 5% CO₂ incubator for 1-4 hours.



Note 1: For adherent cells, gently lift the cells with 0.5 mM EDTA to keep the cells intact, and wash the cells once with serum-containing media prior to incubation with TF2-VAD-FMK.

Note 2: The appropriate incubation time depends on the individual cell type and the cell concentration used. Optimize the incubation time for each experiment.

4. Wash and spin the cells twice. Resuspend the cells in 0.5 ml of assay buffer or growth medium.
Note: TF2-VAD-FMK is fluorescent, thus it is important to wash out any unbound reagent to remove the background.
5. If desired, label the cells with a DNA stain (such as propidium iodide or 7-AAD for dead cells).
6. If desired, fix cells.
7. Monitor the fluorescence intensity with a flow cytometer using the FL1 channel (Ex/Em = 490/525 nm). Gate on the cells of interest, excluding debris.

Data Analysis

In live non-apoptotic cells, TF2-VAD-FMK detects innate apoptosis in non-induced cells, which is typically 2-6% of all cells. In apoptotic cells, TF2-VAD-FMK binds to active caspases resulted in increased staining intensity.

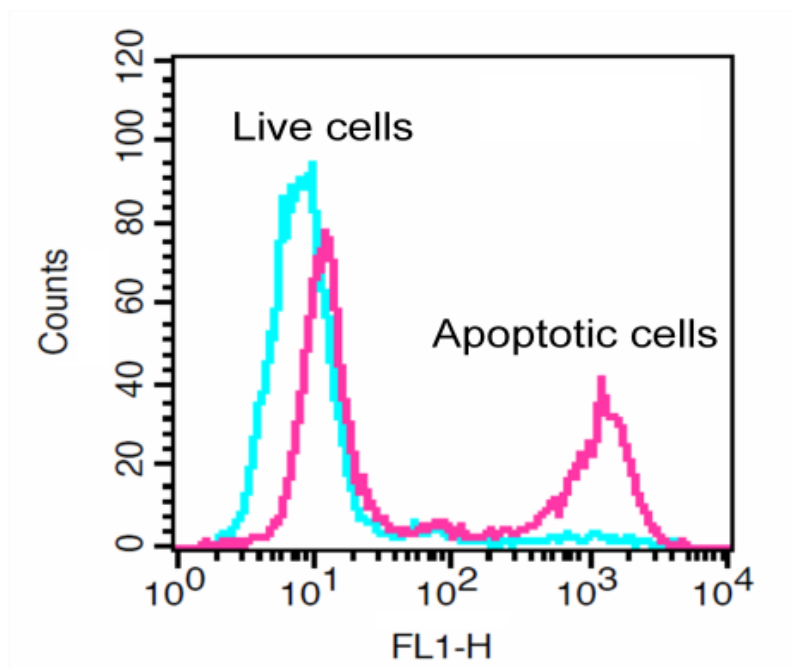


Figure 1. The increase in TF2-VAD-FMK fluorescence intensity with the addition of Camptothecin in Jurkat cells. Jurkat cells were treated without (Blue) or with 20 μ M camptothecin (Red) in a 37 $^{\circ}$ C, 5% CO₂ incubator for 4-5 hours, then dye loaded with TF2-VAD-FMK for 1 hour.

References

1. Li JN, Song DQ, Jiang JD. (2004) [Antitumor mechanism of 3-bromopropionylamino benzoylurea on leukemia and lymphoma]. Yao Xue Xue Bao, 39, 491.
2. Thrane C, Kaufmann U, Stummann BM, Olsson S. (2004) Activation of caspase-like activity and poly (ADP-ribose) polymerase degradation during sporulation in *Aspergillus nidulans*. Fungal Genet Biol, 41, 361.
3. Pandey S, Smith B, Walker PR, Sikorska M. (2000) Caspase-dependent and independent cell death in rat hepatoma 5123tc cells. Apoptosis, 5, 265.

